



ISSN Print: 2394-7500
 ISSN Online: 2394-5869
 Impact Factor: 5.2
 IJAR 2018; 4(3): 231-233
 www.allresearchjournal.com
 Received: 01-01-2018
 Accepted: 02-02-2018

Achaiah Nalamada
 Dept of Zoology, Kakatiya
 University, Warangal
 Telangana, India

N Vijaya Kumar
 Dept of Zoology, Kakatiya
 University, Warangal,
 Telangana, India.

A study on the protein content of cyclophyllidean cestode, *Raillietina tetragona* (Molin, 1858) infecting domestic chick, *Gallus gallus*

Achaiah Nalamada and N Vijaya Kumar

Abstract

Raillietina tetragona is an endogenous helminth parasite infecting domestic chick. In the present study, total proteins, soluble and insoluble proteins levels in immature, mature and gravid proglottids of *Raillietina tetragona* have been undertaken. The total proteins in immature, mature and gravid proglottids are 96.42 ± 1.8 , 90.37 ± 2.1 and 62.16 ± 1.1 mg protein/g fresh weight respectively. Soluble protein levels are 48.95 ± 1.9 , 46.57 ± 1.0 , 42.23 ± 1.5 and insoluble protein levels are 52.37 ± 1.5 , 39.43 ± 1.0 , and 35.85 ± 1.9 mg protein /g fresh weight respectively. The data obtained from immature, mature and gravid segments of adult worm supports the transition in proteins levels in relation to the differentiation of proglottids. The reason for higher protein content in immature region indicates rapid proliferation rate, as it is involved in strobilisation which in turn requires amplified metabolic activity. The same was observed for soluble and insoluble proteins too and the data was statistically analysed by ANOVA.

Keywords: Differentiation of proglottids, *Raillietina tetragona*, total proteins, soluble and insoluble proteins, ANOVA

Introduction

Cestodes are parasitic helminths. They occupy alimentary canal. Livestock in the tropics generates substantial revenue, provides employment, supplies manure and animal protein. Their health status and productivity to fulfill these multipurpose functions is sometimes sub-optimal because of diseases. Helminthosis remains one of the world's most prevalent and economically significant parasitosis of man and domestic animals. Gastro-intestinal cestodes have been identified as one of the causes for production losses which arise primarily through mortality, severe weight loss, decreased egg and meat production, impaired metabolic and reproductive activities.

Cestodes depend on their hosts for energy sources ^[1]. Carbohydrates are most commonly used source of energy in cestodes ^[2]. Glycogen is the major energy source under both aerobic and anaerobic conditions ^[3]. However, large reserves of proteins were detected in cestode worms.

Several workers reported the protein levels from the whole worm, in the present study an attempt was made to study the differential distribution of protein in different proglottids of the adult worm. It has been reported that protein content was reported as lower than that of the invertebrates.

In parasitic helminths, the protein usually constitutes between 20-40% of the dry weight ^[4-5], over 60% have been reported for *Echinococcus spp* ^[6]. In some cases, it is as high as 70% of the dry weight, reported from *Macracanthorhynchus hirudinaceus*, and the infective larvae of *Nippostrongylus brasiliensis* ^[7] and 32% of protein content was reported from *Hymenolepis diminuta*. Differential distribution of protein in immature, mature and gravid regions of cestode parasites *Avitellina lahorea* and *Styletia globipunctata* was reported ^[8]. Some quantitative studies on proteins of some cestodes collected from different hosts and localities of Western Maharashtra were made ^[9]. Several workers reported protein levels from different helminth parasites infecting different hosts ^[10-15].

Correspondence
Achaiah Nalamada
 Dept of Zoology, Kakatiya
 University, Warangal,
 Telangana, India

The material and methods

Raillietina tetragona is a poultry cestode, infects domestic fowl *Gallus gallus*. Worms were collected from viscera of naturally infected birds from freshly slaughtered chicks at Warangal abattoir. The intestine was dissected and parasites were collected and washed with 0.9% normal saline and washed in several changes to remove debris, mucus etc. Identification was carried out with the help of Systema Helminthum^[16] and Helminths, arthropods and protozoa of domesticated animals^[17]. The protein content in the tissue was estimated by Folin-phenol method as described by Lowry *et al*^[18].

Results and Discussion

The total proteins, soluble and insoluble protein values in *Raillietina tetragona* are summarized in table-1 and histogram-1 respectively. The total proteins in immature, mature and gravid proglottids are 96.42 ± 1.8 , 90.37 ± 2.1 and 62.16 ± 1.1 mg protein/g fresh weight respectively. Soluble protein levels are 48.95 ± 1.9 , 46.57 ± 1.0 , 42.23 ± 1.5 and insoluble protein levels are 52.37 ± 1.5 , 39.43 ± 1.0 , and 35.85 ± 1.9 mg protein /g fresh weight respectively.

Table no.1 showing the levels of different protein fractions (Each value is a mean of six values with \pm SE)

Content	Immature	Mature	Gravid
Total proteins	96.42 ± 1.8	90.37 ± 2.1	62.16 ± 1.1
Soluble proteins	48.95 ± 1.9	39.43 ± 1.0	42.23 ± 1.5
Insoluble proteins	52.37 ± 1.5	39.43 ± 1.0	35.85 ± 1.9

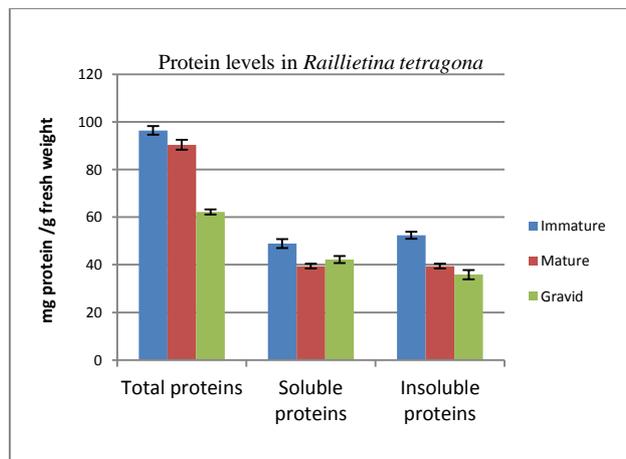


Fig 1: showing the levels of different protein fractions.

There exists a difference in the distribution of protein along the strobilar region, with the highest value in immature followed by mature and gravid regions. The mature and gravid regions had about 6% and 2% less protein content in relation to that of immature region. The values of total proteins for *Raillietina tetragona* are comparable with the previous reports. The decrease in protein content is in agreement with the findings of from *Hymenolepis diminuta*^[19].

A total of 32% protein content was reported in *Hymenolepis diminuta*^[20] and some authors reported higher content of protein in mature proglottids of *Hymenolepis diminuta*^[21]. Fast growth of cestodes needs a high rate of synthesis of proteins. Protein synthesis was highest in immature proglottids^[22]. The total proteins were studied by PAGE and iso-enzyme pattern and it is found that in all parts it is similar^[23].

In *Cittotenia perplexa* 21% of protein, 22% from *Moneizia expansa*^[24], 61% from *Echinococcus granulosus*^[25], 36% from *Raillietina cesticillus*^[26] and from *Taenia taenaeformis* larva it was 29% and in adults it was 45%^[27]. Protein content depends on age, degree of maturation and previous metabolic history of organism. There are two protein fractions viz., soluble and insoluble. Soluble proteins include components like enzymes, glycoproteins, lipoproteins and antigens while insoluble proteins include keratin, collagen and sclerotin. Proteins of soluble and insoluble nature have been reported in the tissues of some trematodes, nematodes and in hooks of some cestodes scolices^[28]. Sclero and chromo proteins found in digeneans and pseudophylledean cestodes^[29]. Chromo protein is the food for many helminth parasites as it contains oxidative pigments like haemoglobin. The highest content in immature region may be due to its proliferation rate which requires more protein. The reason for higher protein content in immature region indicates rapid proliferation rate, as it is involved in strobilisation which requires amplified metabolic activity. The same was observed for soluble and insoluble proteins.

Acknowledgements

The author acknowledges the financial support of Kakatiya University in the form of University Research Fellowship during the research period (2009-2013).

References

- Smyth, University Reviews in Biology. Oliver and Boyd Ltd. Edinburg. 1969; 11.
- Cheng TC. General Parasitology. by academic Press, Inc Orlando, florida. 1986; 2.
- Agosin and Repetto Studies on the metabolism of *Echinococcus granulosus*. IX. Protein synthesis in scolices. Experimental Parasitology. 1967; 21:195-208.
- Smyth. University Reviews in Biology. Oliver and Boyd Ltd. Edinburg. 1969; 11.
- John Barrett. Text book of "Biochemistry of parasitic helminths". University Park Press, Baltimore, US. 1981.
- Barbara M MacKinnon, Michael DB. Burt Histological and ultra structural observations on the secondary scolex and strobila of *Haplobothrium globuliforme* (Cestoda: Haplobothrioidea) Canadian Journal of Zoology. 1985; 63(8):1995-2000.
- Goodchild. Protein contents of the tapeworm *Hymenolepis diminuta* from normal, bile-less, and starved rats. J. Parasit. 1961; 47:830-832.
- Rao Patwari Raghavendra. Studies on some biochemical and physiological aspects of two cestodes. Ph.D thesis, Osmania University, India. 1982.
- Hiware *et al*. Quantitative studies on proteins in some cestodes collected from different hosts and localities of western Maharashtra. Society India. 2002; 152-156.
- Dhongde *et al*. protein profiles of avian cestode A case study-the biosphere. International journal of lifesciences. 2010; 2:133-136.
- Radfar *et al*. Hematological, biochemical and pathological findings in goats naturally infection with *Cysticercus tenuicollis*, J.of.Parasitic Diseases, 2012. DOI 10.1007/s 12639-012-0188-y.
- Humbe *et al*. Biochemical studies of gastrointestinal cestode parasites in *Capra hircus* and *Ovis bharal* from

- Marathwada region. *Int. Multidiscip Res. J.* 2011; 12:08-10.
13. Jawale *et al.* Biochemical studies of Caryophyllidean tapeworms in fresh water fish *Clarius batrachus*. *Recent. Res. Sci. Technol.* 2011; 3(3):35-36.
 14. Jawale, Fartade, Borde Biochemical studies of Caryophyllidean tapeworms in fresh water fish *Clarias batrachus*. *Recent Res Sci Technol.* 2011; 3(3):35-36.
 15. Debraj Biswal. Biochemical and molecular characterization of the cyclophyllidean cestode, *Cotugnia cuneata* (Meggit, 1924), an endoparasite of domestic pigeons, *Columba livia domestica*. 2012.
 16. Yamaguti S. *Systema Helminthum. Cestodes of Vertebrates.* Interscience Publishers Inc. New York. 1959; 2.
 17. Soulsby E.J.L. *Helminths, Arthropods and protozoa of domesticated animals*, London Bailliere, Tindall. 1986; 7:809.
 18. Lowry OH, Rosenbrough NJ, Farr AL, Randall RJ. The method for protein estimation. *J Biol Chem.* 1951; 193:265.
 19. Fairbairn *et al.* biochemistry of normal and irradiated strains of *Hymenolepis diminuta*. *Experimental Parasitology.* 1961; 11:248-263.
 20. Goodchild *et al.* The growth of rat tapeworm, *Hymenolepis diminuta* during the first five days in final host *J of Parasit.* 1961; 47:819-829.
 21. Fairbairn *et al.* Biochemistry of normal and irradiated strains of *Hymenolepis diminuta*. *Experimental Parasitology.* 1961; 11:248-263.
 22. Roberties *et al.* *Comp. Biochem. Physiology.* 1971; 40A:777-787.
 23. Bursey Mc Kenzie, Burt Polyacrylamide gel electrophoresis in differentiation of *Taenia* (Cestoda) by total protein. *Int J Parasitol.* 1980; 10:167-174.
 24. Campbell. Nitrogen, amino acid composition of three species of anoplocephalid cestodes *Moneizia*, *Thysanosoma* and *Cittotenia*. *Exp. Parasit.* 1960; 9.
 25. Agosin *et al.* Studies on metabolism of *Echinococcus granulosus*. *Exp. parasit.* 1957; 6:37-51.
 26. Reid. Certain nutritional requirements of the fowl cestode *Raillietina cesticillus* (Molin) as demonstrated by short periods of starvation of the host. *The Journal of Parasitology.* 1942; 28:(4)319-340
 27. Von Brand. IBR Bowman Studies on the aerobic and anaerobic metabolism of larval and adult *Taenia taeniaeformis* - *Experimental Parasitology.* 1961; 11:276-297.
 28. Gallagher. Chemical composition of hooks isolated from hydatid scolices IHC-*Experimental Parasitology.* 1964; 110-117.
 29. Smyth, Clegg Egg-shell formation in trematodes and cestodes. *Exp Parasitol.* 1959; 8(3):286-323.