International Journal of Applied Research 2018; 4(9): 01-04



International Journal of Applied Research

ISSN Print: 2394-7500 ISSN Online: 2394-5869 Impact Factor: 5.2 IJAR 2018; 4(9): 01-04 www.allresearchjournal.com Received: 01-07-2018 Accepted: 03-08-2018

Dr. Anil S Joshi

Associate Professor, Dept. of Medicine: Govt. Medical College and Hospital, Aurangabad, Maharashtra, India

Sayali S Shirkhedkar

Govt. College of Pharmacy, Opposite Govt. Polytechnic, Osmanpura, Aurangabad, Maharashtra, India

Dr. SR Shahi

Assistant Professor, Govt. College of Pharmacy, Aurangabad, Maharashtra, India

Rishik Sharma

Govt. College of Pharmacy Aurangabad, Maharashtra, India

Shivam Tiwari

Govt. College of Pharmacy Aurangabad, Maharashtra, India

Aishwarya Khadke

Govt. College of Pharmacy Aurangabad, Maharashtra, India

Preeti Naikwade

Govt. College of Pharmacy Aurangabad, Maharashtra, India

Correspondence Sayali S Shirkhedkar Govt. College of Pharmacy, Opposite Govt. Polytechnic, Osmanpura, Aurangabad, Maharashtra, India

Study antibiotic sensitivity and resistance pattern from sputum / tracheal swab of patients on ventilatory support in ICU of tertiary care hospital

Dr. Anil S Joshi, Sayali S Shirkhedkar, Dr. SR Shahi, Rishik Sharma, Shivam Tiwari, Aishwarya Khadke and Preeti Naikwade

Abstract

Objective: To study antibiotic sensitivity and resistance pattern from sputum/tracheal swab of patients on ventilatory support in ICU of tertiary care hospital.

Method: A prospective observational study was carried out based upon the reports of bacterial isolates from ICU of Dept. of Medicine, Govt. Medical College and Hospital, Aurangabad between "23 October, 2017 to 28 February, 2018". 56 patients were studied. Susceptibility testing was performed using disk diffusion method. The results were interpreted according to the guidelines of the Clinical and Laboratory Standards Institute (CLSI).

Results: The study revealed the presence of Klebsiella species in 28.29% (n=26) samples followed by *Pseudomonas aeruginosa* 20.69% (n=18), Klebsiella Pneumonia 13.79% (n=12), Acinetobacter baumannii 10.34% (n=9), Non-fermenting gram negative rods 9.19% (n=8), respectively. A high range of resistance was seen for Ceftazidime in case of Klebsiella spp (91.66%). Overall Colistin was the most sensitive drug among all group of microorganisms.

Conclusion: The present study is an initiative to observe serious threats in patients admitted in ICU due to nosocomial infection. Most com monest microorganisms observed were *Klebsiella* species 28.29% (n=26), *Pseudomonas aeruginosa* 20.69% (n=18), *Klebsiella pneumonia* 13.79% (n=12), *Acinetobacter baumanni* 10.34% (n=9). Maximum resistance was observed to Ceftazidime, 3rd generation cephalosporin 92.7% (51 out of 55). Highest sensitivity was seen for Colistin (80%) followed by Ciprofloxacin (76.8%). Such studies will restrict emergence of resistant bacteria, effective cost benefit, inventory and stock management.

Keywords: Antimicrobial resistance, multidrug resistance, antibiotic sensitivity

Introduction

Amongst the healthcare settings, intensive care unit (ICU) is known to be "Canopy of infections". Not only the invasive procedures but also drugs increases the risk of infection. To understand and overcome this on-going situation, the sensitivity and resistance pattern studies need to be conducted [1] it is being observed through various studies that 20-30% of ICU admissions had nosocomial infection. The multicentre trials comprising 1265 ICU's all over 75 countries revealed that the hospital acquired infection was present in 50% of ICU patients [2] This may be accounted due to many invasive procedures are being carried out in the ICUs. Certain therapeutic interventions responsible for infectious complications includes indwelling catheters, sophisticated life support, intravenous fluid therapy, prosthetic devices, immunosuppressive therapy and use of broad spectrum antibiotics leading to a spectrum of multi-drug resistant pathogens. These factors contribute for emergence of nosocomial infections [3] the organisms causing infections and their antibiotic resistance patterns vary throughout the hospital settings.

The International study of infections in ICU, which was conducted in 2007, demonstrated that the patients who had longer ICU stays had higher rates of infection, especially infections due to resistant *Staphylococci*, *Acinetobacter*, *Pseudomonas* species, *Candida* species ^[7] In an another study on bacterial isolates from in-patients and out-patient attendees at the Eric Williams Medical Sciences complex, a 560-bed medical complex located in the north western part of Trinidad, represented about 2.74% (31 out of 1129) of all culture positive

isolates from clinical sources ^[8] *Pseudomonas aeruginosa* and *Enterobacteriaceae* species are the major cause of Healthcare associated infections (HAI's) often with significant drug resistance leading to increased morbidity and mortality. Approximately, 2-10% of *P. aeruginosa* are resistant to all available treatments ^[7]

EPIC II study showed that 51% of patients were considered to be infected while admitted in ICU. The infection was of respiratory origin in 64% of cases. *S. aureus* (20.5%) was the most frequent organism isolated, despite the overall predominance of Gram-negative organisms as a group: 62.2% (*E. coli, Enterobacter* spp., *Klebsiella* spp., *Pseudomonas* spp. and *Acinetobacter* spp.) [9]

The aim of the present research is to study the antibiotic resistance and sensitivity patterns for patients admitted in ICU Department of Medicine of Government Medical College and Hospital, Aurangabad.

Materials and Method

A prospective observational study was carried out based upon the reports of bacterial isolates from ICU of Department of Medicine, Govt. Medical College and Hospital, Aurangabad with 16 beds between the periods "23 October, 2017 to 28 February, 2018". Out of all patients admitted, 56 patients were studied.

The approval from Institutional Ethics Committee was granted and approved on date 23 October 2017 with No. Pharma/IEC-GMCA/518/2017. Individual patient's

informed consent was taken since the patient file was taken for collecting the diagnostics and therapeutic data. Susceptibility testing was performed using disk diffusion method. The results were interpreted according to the guidelines of the Clinical and Laboratory Standards Institute (CLSI)

Results

For the present study the mean age group was between 30-39 years in which 58.92% were males and 41.08% females. *Klebsiella* species was seen in 28.29% (n=26) samples, followed by *Pseudomonas aeruginosa* 20.69% (n=18), *Klebsiella Pneumonia* 13.79% (n=12), *Acinetobacter baumanni* 10.34% (n=9), Non-fermenting *gram negative rods* 9.19% (n=8), *Acinetobacter spp* 3.45% (n=3), Non-*Fragmenting gram negative rods*, Non-pigmented *gram negative rods* and *E. Coli* 3.57% (n=2), *Staphylococcus aureus*, Non-pigmented *Pseudomonas aeruginosa*, *Acinetobacter iwoffi*, and *proteus spp* had an incidence of 1.14% (n=1). Table 1 shows the distribution of various microorganisms.

For *Klebsiella spp*, ciprofloxacin 30% (n=6) was found to be the most sensitive antibiotic and Ceftazidime 91.66% (n=22) was the most resistant antibiotic. Similar pattern was observed in *Pseudomonas aeruginosa*. Maximum resistance was observed for Ceftazidime, 3rd generation cephalosporin 92.7% (51 out of 55). Colistin was found to be the most sensitive drug with overall sensitivity 80% (8/10).

Microorganisms	Frequency	Percentage (%)
Klebsiella. Spp.	26	28.29
Pseudomonas aeruginosa	18	20.69
Klebsiella. Pneumonia	12	13.79
Acinetobacter baumanni	9	10.34
Non-fermenting gram-ve rods	8	9.19
Acinetobacter species	3	3.45
Non-pigmented Pseudomonas aeruginosa	2	2.29
Non-fragmenting gram-ve rods	2	2.29
Escherichia coli	2	2.29
Staphylococcus aureus	1	1.14
Acinetobacter iwoffi	1	1.14
Non-pigmented gram negative rods	1	1.14
Proteus spp.	1	1.14

Table 1: Frequency and percentage of bacterial species

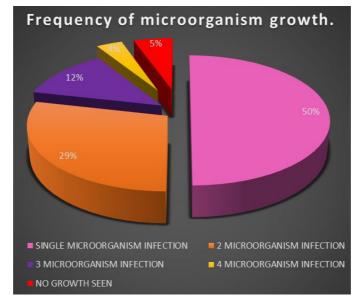


Fig 1: Frequency of microorganism growth

Table 2: Antibiotic susceptibility pattern

Antibiotic	Frequency of Test	Name of Organism	Frequency of tested in organism	Sensitivity (%) (n = Samples found)
Ceftazidime	78	Klebsiella. spp	23	4.34% (n=1)
		Pseudomonas aeruginosa	16	37.5% (n=6)
		Klebsiella pneumonia	7	0% (n=0)
		Acinetobacter baumanni	9	22.22% (n=2)
Ciprofloxacin	82	Klebsiella. spp	26	23.07% (n=6)
		Pseudomonas aeruginosa	16	43.75% (n=7)
		Klebsiella pneumonia	12	25% (n=3)
		Acinetobacter baumanni	9	22.22% (n=2)
Piperacillin + Tazobactum	53	Klebsiella spp	15	20% (n=3)
		Pseudomonas aeruginosa	9	33.33% (n=3)
		Klebsiella pneumonia	8	25% (n=2)
		Acinetobacter baumanni	5	40% (n=2)
	46	Klebsiella spp	18	16.66% (n=3)
		Pseudomonas aeruginosa	10	20% (n=2)
Ceftazidime + Clavulanic acid		Klebsiella pneumonia	3	0% (n=0)
		Acinetobacter baumanni	4	0% (n=0)
	46	Klebsiella spp	17	17.64% (n=3)
		Pseudomonas aeruginosa	8	37.5% (n=3)
Amikacin		Klebsiella. pneumonia	3	0% (n=0)
		Acinetobacter baumanni	3	0%(n=0)
Cefotaxime	37	Klebsiella spp	13	7.69% (n=1)
		Pseudomonas aeruginosa	6	33.33% (n=2)
		Klebsiella pneumonia	7	28.57% (n=2)
		Acinetobacter baumanni	7	14.28% (n=1)
Ceftriaxone	30	Klebsiella spp	8	0% (n=0)
		Pseudomonas aeruginosa	8	37.5% (n=3)
		Klebsiella pneumonia	9	0%(n=0)
		Acinetobacter baumanni	7	14.28% (n=1)
Meropenem	28	Klebsiella spp	11	27.27% (n=3)
		Pseudomonas aeruginosa	4	50% (n=2)
		Klebsiella pneumonia	5	60% (n=3)
		Acinetobacter baumanni	5	40% (n=2)
Cefaperazone	24	Klebsiella spp	9	0% (n=0)
		Pseudomonas aeruginosa	5	60% (n=3)
		Klebsiella pneumonia	5	0% (n=0)
		Acinetobacter baumanni	5	40% (n=2)
Colistin	10	Klebsiella spp	1	100% (n=1)
		Pseudomonas aeruginosa	6	83.33% (n=5)
		Klebsiella pneumonia	1	100% (n=1)
		Acinetobacter baumanni	1	100% (n=1)

Discussion

The study revealed the presence of Klebsiella species in 28.29% (n=26) samples followed by Pseudomonas aeruginosa 20.69% (n=18), Klebsiella Pneumonia 13.79% (n=12), Acinetobacter baumanni 10.34% (n=9), Nonfermenting gram negative rods 9.19% (n=8), Acinetobacter spp 3.45% (n=3), Non-Fragmenting gram negative rods, Non-pigmented gram negative rods and E. Coli 3.57% (n=2),Staphylococcus aureus, Non-pigmented Pseudomonas aeruginosa, Acinetobacter iwoffi, and proteus spp had an incidence of 1.14% (n=1). Similar results were observed in the study by Mahin Jamshidi et al titled, "Antimicrobial resistance pattern among intensive care unit patients." The study revealed that the most frequent gram negative microorganisms were P. aeruginosa (43.2%) and Klebsiella spp (33.7%); Staphylococcus aureus (39.2%) was most common gram positive microorganism isolated [7].

The frequency for the growth of microorganism was observed as follows: 50% patients (28/56) single microorganism, 28.57% patients (16/56)two microorganisms, 12.50% patients (7/56)three 3.57% microorganism, and patients (2/56)microorganisms, respectively. The study also revealed that 5.35% (3/56) of patients were sterile throughout the stay. Fig.1 shows the distribution for frequency microorganisms growth.

The incidence of single microorganism growth was highest as compared to the multiple microorganisms though very few of them were infectious. This may be accounted to good infection control practices like maintenance of good hygienic conditions, and following proper sampling procedures and other invasive techniques.

A similar pattern was observed in the study entitled, "Nosocomial bacterial infections and their antimicrobial susceptibility patterns among patients in intensive care units: A cross sectional study" by Peter Agaba *et al.* of Uganda. The data revealed that 32 patients developed nosocomial infection and a total of 52 isolates were obtained. About 20 (62.5%) of the patients grew one organism, 11 (34.37%) grew two organisms and only 1 (3.12%) patient grew three organisms [^{23]}.

A high resistance was seen for the drug Ceftazidime, *Klebsiella spp* (91.66%), *Pseudomonas aeruginosa* (62.5%) and *Klebsiella pneumonia* (100%), respectively, while the drug Ciprofloxacin was sensitive. Table 2 shows the detailed distribution of sensitivity and resistance pattern.

The study titled, "Antimicrobial resistance pattern among aerobic gram negative bacilli of lower respiratory tract specimens of Intensive Care Unit patients in a neuro centre" of Bangalore by H.B. Veena Kumari, *et al* reported *Pseudomonas aeruginosa* of tracheal and bronchial specimens showed 59.8% and 40% resistance to Amikacin,

respectively. *Non-fermentative gram-negative bacilli* exhibited 71.4% and 75% resistance to Amikacin and 38.3% and 62.5% to netilmicin in tracheal and bronchial specimens, respectively. Netilmicin showed highest activity against *NFGNB* and Amikacin against *Klebsiella spp* and *Escherichia coli* specimens [17].

Summary and Conclusion

The present study is an initiative to observe serious threats in patients admitted in ICU due to nosocomial infection. The commonest microorganisms observed were *Klebsiella* species 28.29% (n=26), *Pseudomonas aeruginosa* 20.69% (n=18), *Klebsiella pneumonia* 13.79% (n=12), *Acinetobacter baumanni* 10.34% (n=9). Maximum resistance was observed to Ceftazidime, 3rd generation cephalosporin 92.7% (51 out of 55). Highest sensitivity was seen for Colistin 80% (8 out of 10) followed by Ciprofloxacin76.82% (63 out of 82).

In ICU, there is need of regular testing of antibiotics sensitivity patterns, guiding clinicians selecting therapy. This will restrict emergence of resistant bacteria, effective cost benefit, inventory and stock management.

References

- 1. Radji M, Fauziah S, Aribinuko N. Antibiotic Sensitivity Pattern of Bacterial Pathogens in the Intensive Care Unit of Fatmawati Hospital, Indonesia. Asian Pac. J Trop. Biomed. 2011; 1(1):39-42.
- Tran GM, Ho Le TP, Ha DT, Tran-Nguyen CH, Nguyen TSM, Pham TTN et al. Patterns of Antimicrobial Resistance in Intensive Care Unit Patients: A Study in Vietnam. BMC Infect. Dis. 2017; 17(1)
- Sheth KV, Patel TK, Malek SS, Tripathi C. Antibiotic Sensitivity Pattern of Bacterial Isolates from the Intensive Care Unit of a Tertiary Care Hospital in India. J Pharm Res Trop. J Pharm. Res. December Trop J Pharm Res. 2012; 1111(1166):991-991.
- 4. WHO. WHO | Antimicrobial Resistance. Who. 2017.
- 5. WHO. WHO's First Global Report on Antibiotic Resistance Reveals Serious, Worldwide Threat to Public Health. World Health Organization. News Release, 2014.
- Matuschek E, Brown DFJ, Kahlmeter G. Development of the EUCAST Disk Diffusion Antimicrobial Susceptibility Testing Method and Its Implementation in Routine Microbiology Laboratories. Clin. Microbiol. Infect. 2014; 20(4).
- 7. Jamshidi M, Javadpour S, Eftekhari TE, Moradi N, Jomehpour F. Antimicrobial Resistance Pattern among Intensive Care Unit Patients. African J Microbiol. Res. 2009; 3(10):590-594.
- 8. Mohanty S, Singhal R, Sood S, Dhawan B, Kapil A, Das BK. Citrobacter Infections in a Tertiary Care Hospital in Northern India. J Infect. 2007; 54(1):58-64.
- 9. Subbalaxmi MVS, Lakshmi V, Lavanya V. Antibiotic Resistance Experience in a Tertiary Care Hospital in South India. 2010; 58:18-22.
- 10. CLSI. Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Sixth Informational Supplement. CLSI Document, 2016, M100-S26;
- Adnan M, Mohammad S, Sultana R, Mahmud NU, Sanjana K, Imtiaj H. A Cross Sectional Study on Antibiotic Resistance Pattern of Salmonella Typhi

- Clinical Isolates from Bangladesh. Asian Pac. J Trop. Biomed. 2014; 4:306-311.
- 12. George DF, Gbedema SY, Agyare C, Adu F, Boamah VE, Tawiah AA *et al.* Antibiotic Resistance Patterns of Escherichia Coli Isolates from Hospitals in Kumasi, Ghana. ISRN Microbiol. 2012, 658470.
- 13. Kumari HBV, Nagarathna S, Chandramuki A. Antimicrobial Resistance Pattern among Aerobic Gram-Negative Bacilli of Lower Respiratory Tract Specimens of Intensive Care Unit Patients in a Neurocentre. Indian J Chest Dis. Allied Sci. 2007; 49(1):19-22.
- 14. Baditoiu L, Axente C, Lungeanu D, Muntean D, Horhat F, Moldovan R *et al.* Intensive Care Antibiotic Consumption and Resistance Patterns: A Cross-Correlation Analysis. Ann. Clin. Microbiol. Antimicrob. 2017; 16(1).
- Kumar JAJU, Srinivasa H, Chiramal JA. A Comparative Study of Drug Susceptibility Testing Techniques for Identification of Drug Resistant TB in a Tertiary Care Centre, South India. J Tuberc. Res. 2017; 05(01):44-57.
- 16. Khety Z, Mohanta G, Jain S, Dawoodi S. Changing Antimicrobial Resistance Pattern of Isolates from an ICU over a 3 Year Period. J Assoc. Physicians India. 2017; 65(2):13-16.
- 17. Kumari HBV, Nagarathna S, Chandramuki A. Antimicrobial Resistance Pattern among Aerobic Gram-Negative Bacilli of Lower Respiratory Tract Specimens of Intensive Care Unit Patients in a Neuro centre. Indian J Chest Dis. Allied Sci. 2007; 49:19-22.
- 18. Kumarasamy KK, Toleman MA, Walsh TR, Bagaria J, Butt F, Balakrishnan R *et al.* Emergence of a New Antibiotic Resistance Mechanism in India, Pakistan, and the UK: A Molecular, Biological, and Epidemiological Study. Lancet Infect. Dis. 2010; 10(9):597-602.
- 19. Clark NM, Patterson J, Lynch JP. Antimicrobial Resistance among Gram-Negative Organisms in the Intensive Care Unit. Current Opinion in Critical Care. 2003, 413-423.
- 20. Samah G, Hatem MES, El KAT, Nikhat M. Antimicrobial Resistance Patterns of Klebsiella Isolates from Clinical Samples in a Saudi Hospital. African J Microbiol. Res. 2017; 11(23):965-971.
- 21. Saravanan R, Raveendaran V. Antimicrobial Resistance Pattern in a Tertiary Care Hospital: An Observational Study. J Basic Clin. Pharm. 2013; 4(3):56.
- 22. Mohanty S, Singhal R, Sood S, Dhawan B, Kapil A, Das BK. Citrobacter Infections in a Tertiary Care Hospital in Northern India. J Infect. 2007; 54(1):58-64.
- 23. Agaba P, Tumukunde J, Tindimwebwa JVB, Kwizera A. Nosocomial Bacterial Infections and Their Antimicrobial Susceptibility Patterns among Patients in Ugandan Intensive Care Units: A Cross Sectional Study. 2017; (10):349.
- 24. Mannan A, Shohel M, Rajia S, Mahmud NU, Kabir S, Hasan I. A Cross Sectional Study on Antibiotic Resistance Pattern of Salmonella Typhi Clinical Isolates from Bangladesh. Asian Pac. J Trop. Biomed. 2014; 4(4):306-311.